

Workflows management: new abilities for the biological information overflow

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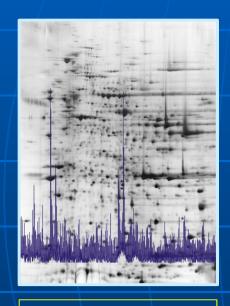
CNR - National Research Council ITB - Institute of Biomedical Technologies

Workflows management: new abilities for the biological information overflow

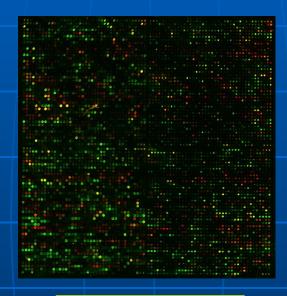
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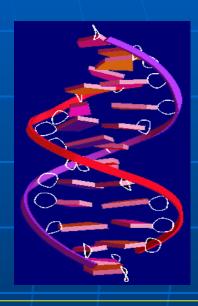
Complex Disease Mapping



Proteins (Proteome)



Microarray (Genome)

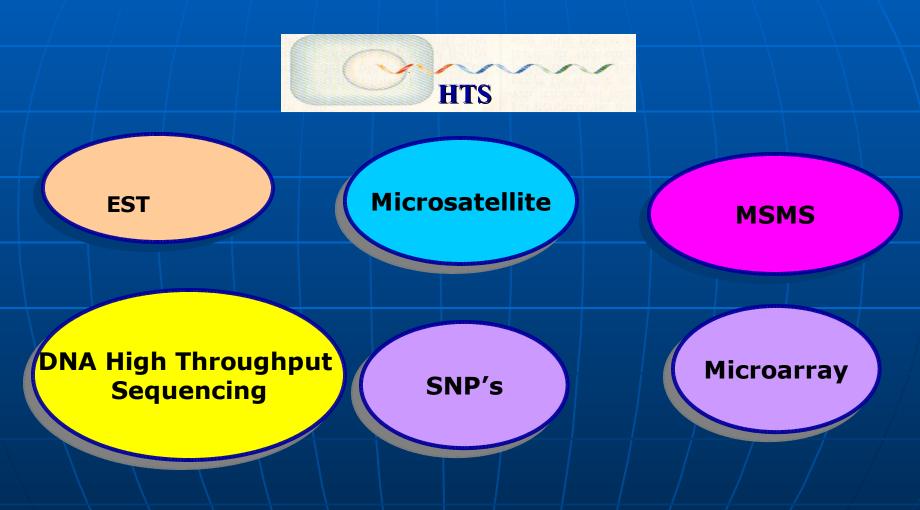


Gene & SNPs (Genome)

Bioinformatics: Emerging Opportunities and Emerging Gaps

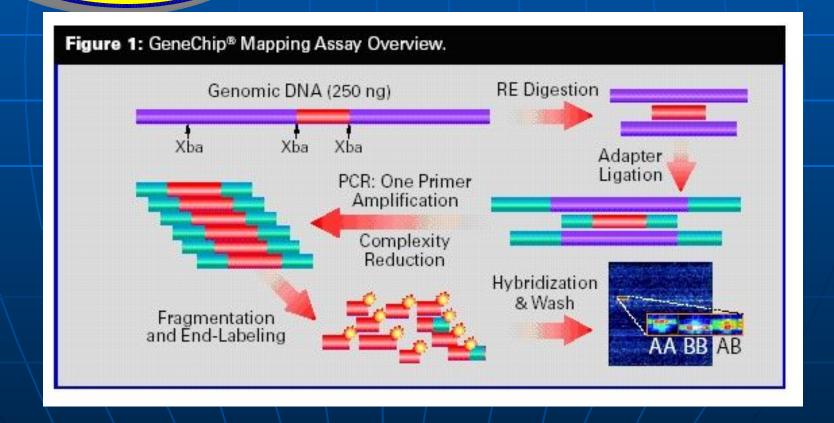
Paula E.Stephan and Grant Black

- A typical gene lab can produce <u>100 terabytes of</u> <u>information a year</u>, the equivalent of 1 million encyclopedias.
- Few biologists have the computational skills needed to fully explore such an astonishing amount of data; nor do they have the skills to explore the exploding amount of data being generated from clinical trials.
- The immense amount of data that are available, and the knowledge that this is but the tip of the data iceberg.



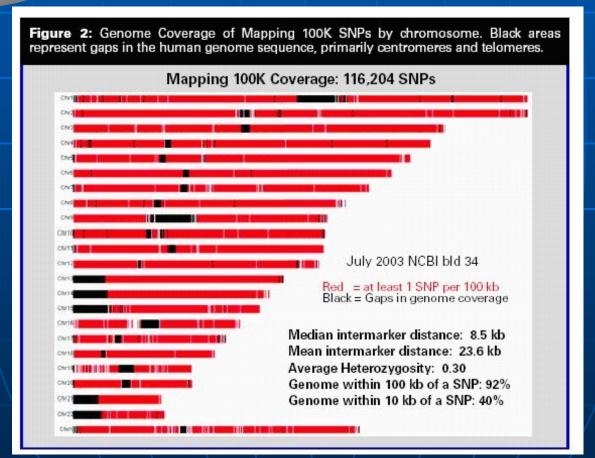
DNA High Throughput
Sequencing





SNP's

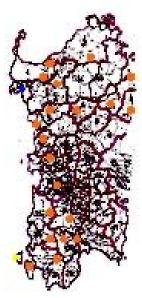




SNP's



The Sardinian Challenge



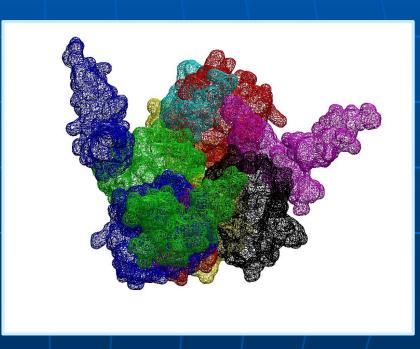
From homologies of surnames, linguistic roots and genetic markers, Sardinians have been subdivided into 31 subpopulations. Long term isolated villages within these subpopulations can be considered as true "Mendelian Breeding Units (MBUs)" just like the precious collection of the Jackson Lab (N. H.) highly inbred mouse strains whose contribution to the study of mammalian genetics (Homo sapiens included) has been, and still is, simply outstanding.



ANDTIASQDT PAKI VII ANK LKI LKDYVDD LKTYNNTYSN VVTVAGEDRI ETAIELSSKY YNSDDKNAIT DKAVNDIVLV GSTSIVDGLV ASPLASEKTA PLLLTSK DKL DSSVKSEIKR VMNLKSDTGI NTSKKVYLAG GVNSISKDVE NELKNMGLKV TRLSGEDRYE TSLAIADEIG LDNDKAFVVG GTGLADAMSI APVASQLKDG DATPIVVVDG KAKEISDDAK SFLGTSDVDI IGGKNSVSKE IEESIDSATG KTPDRISGDD RQATNAEVLK EDDYFTDGEV VNYFVAK DGS TKEDQLVDAL AAAPIAGRFK ESPAPIILAT DTLSSDQNVAVSK AVPKDGG TNLVQVGKGI ASSVINKMKD LLDM

>monoisotopic mass = **39480**

position sequence (NCBI BLAST link) 18- 22 ANKLK 245- 249 NSVSK 262- 265 TPDR 116- 119 SEIK 344- 347 AVPK 298- 302 DGSTK 266- 271 ISGDDR 163- 168 LSGEDR 116- 120 SEIKR 60- 66 YYNSDDK 224- 230 EISDDAK 155- 159 NMGLK 67- 72 NAITDK 126- 134 SDTGINTSK 160-168 VTRLSGEDR 21- 25 LKDLK 108- 115 DKLDSSVK 135- 147 KVILAGGVNSISI 17 WIE 14-1- 13 ANDTIASQDTPAK 126- 147 SDTGINTSKKVYLAGGVNSISK 14- 20 VVIKANK



Disease Gene

- Mutations in target sequences are usually revealed by either phenotypic selection in experimental test systems or, in case of disease-causing genes in humans, by clinical studies in which certain genes are sequenced in groups of patients and in control groups.
- Both the experimental test systems and the clinical studies rely on detectable (mutable) positions, which are sites where DNA sequence changes cause phenotypic changes.

Human genetic identity

- Genomic sequence 99.9% identical
- 3,200,000 nucleotides different
- Single base differences in genomes between any two individuals: ca. 3 million
- Amino acid differences in proteomes between any two individuals: ca. 100,000

Variation types

- Macro:
 - Chromosome numbers
 - Segmental duplications, rearrangements, and deletions
- Medium:
 - Sequence Repeats
 - Transposable Elements
 - Short Deletions, Sequence and Tandem Repeats (including microsatellites)
- Micro:
 - Single Nucleotide Polymorphisms (SNPs)
 - Single Nucleotide Insertions and Deletions (Indels)

What are Single Nucleotide Polymorphisms (SNPs)?

SNPs result from replication errors and DNA damage

Are all SNPs really SNPs?

- A SNP is found by aligning overlapping DNA sequences and identifying variable positions
- Two types of errors in SNP finding
 - Inclusion of paralogs in sequence alignment
 - Sequencing errors

GCATGCAAGCAGATA
GCATGCAAGCAGATA
GCATGCAAGCAGATA
GCATGCAAGCAGATA
GCATGCAAGCAGATA
GCATGCAAGCAGATA
GCATGCAAGCAGATA
ACCAGCAGATA
ACCAGCAGATA
Naples

Application of SNP data

- Study of evolution
 - Traces evolutionary history of different populations
- DNA fingerprinting
 - criminal or parental verification
- Markers for mapping of polygenic traits
- Genotype-specific medication
- Most genes contain SNPs
 - 93% genes have one or more SNPs
 - 39% have more than 10 SNPs)

Types of SNPs

Genic, coding SNPs

non-synonymous

Maintaining vs. altering protein structure/function synonymous

Maintaining vs. altering splicing

Genic, non-coding SNPs

Regulatory SNPs

Maintaining vs. altering gene expression

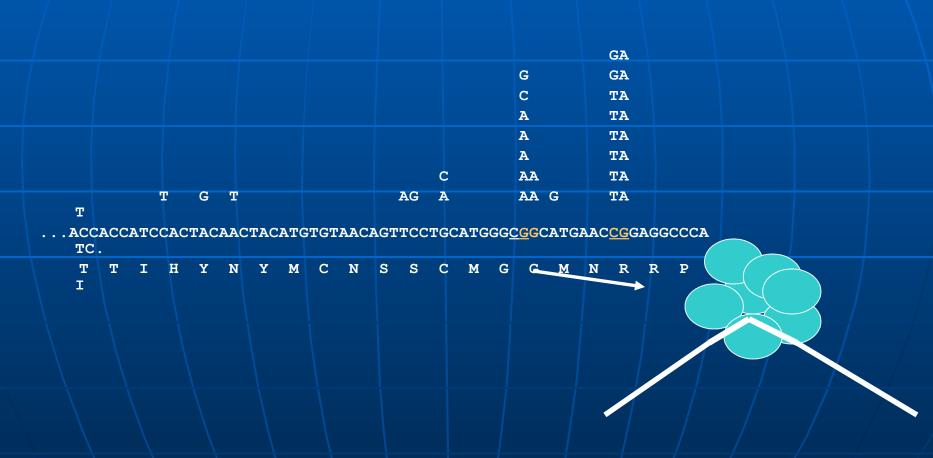
Intronic SNPs

Maintaining vs. altering gene expression/splicing

Linked SNPs

usually intergenic

Context-dependence of mutations



• A mutation spectrum is a distribution of mutation frequencies along nucleotide sequences and is compiled by the analysis of a large number of mutant target sequences.

					Т			AAAA									
	TT				Т			TTTT									
	AT				G			A									
	CA		A		Т		T	A									
	AT		G	G	T		G	A		T							
	AT		AG	GC	A		С	AT		Т							
\T	AG	C	AG	GT	AT		T	TT		A	A						
G	CT	G	CG	AT	AT 7	T	G	GAT	GGG	AG	A C		7		G		
CG	TCT	CC	AT	AT	T CT 7	' \T	GT	TTGAT	CAT	T AT	A AG	Т	<u></u>	С	/ G	G	
ATGT	TCAT	A ACC	TCTC	AC .	A TAT A	A AG	GGG	GTTAG C	CATGAC	TTT TT A	A T CT I	GT	т т	CT	GCG 5	rg c	
GA TA	ГСА <u>GC</u> Т	GA <u>TA</u> TO	CCA <u>GC</u> T	GGA <u>TA</u> T	CAC A<u>GC</u>T (SAGA	TA TCA	aca <u>gc</u> tgaa	GA <u>TA</u> TCA	CACA <u>GC</u> TG	ACAGA <u>TA</u> TO	CACCACA	<u>GC</u> TG	ACCA	GA <u>TA</u> T(CAGTT	100
				7		1											

 Mutation frequencies vary significantly along nucleotide sequences such that mutations often concentrate at certain positions called mutation

Mutation hotspots are frequently caused by mutable motifs.

```
RGYW/WRCY (R = A/G, Y = T/C, W = A/T) G/C mutator WA/TW (hotspot positions are underlined) A/T mutator
```

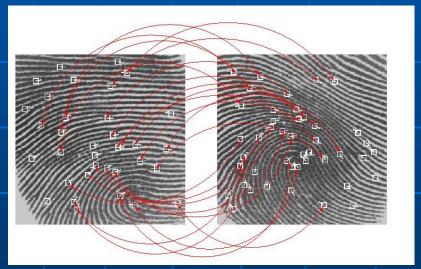
		Т		AAAA						
TT		Т		TTTT						
AT		G		A						
CA	A	Т	Т	A						
AT	G	G T	G	A		T				
AT	AG	GC A	С	AT		T				
T AG	C AG	GT AT	Т	TT		A	А			
G CT	G CG	AT AT	T T G	GAT	GGG	AG	A C	T	G	
CG TCT	CC AT	AT T CT	T T G T	TTGAT	CAT	T AT	A AG	т т	C G G	
ATGT TCAT A	ACC TCTC	AC A TAT	A AG GGG	GTTAG (CATGACT	TT TT A A	T CT T	GT TT	CT GCG TG C	
GA <u>TA</u> TCA GC TGA	TA TCCA GC TGG	A <u>ta</u> tcaca <u>gc</u>	i gaga <u>ta</u> tc <i>i</i>	AACA <u>GC</u> TGAA	AGA <u>TA</u> TCAC	ACA <u>GC</u> TGAC	AGA <u>TA</u> TCA	CCACA <mark>GC</mark> TO	GACCAGA <u>TA</u> TCAGTT	100
W <u>A</u> R <u>G</u> YW	W <u>A</u>									

Hotspot positions in RGYW/WRCY and WA/TW mutable motifs

 $\underline{\mathbf{T}}\mathbf{W}$

WR<u>C</u>Y

Mutation hotspots in DNA reflect intrinsic properties of the mutation process, such as sequence specificity, that manifests itself at the level of interaction between mutagens, DNA, and the action of the repair and replication machineries.



The nucleotide sequence context of mutation hotspots is <u>a fingerprint</u> of interactions between DNA and repair/replication/modification enzymes, and <u>the analysis of hotspot context provides evidence of such interactions</u>.

Context factors may influence mutation rates:

- Homonucleotide runs and microsatellites
- Direct and inverted repeats
- Local mononucleotide composition
- DNA conformation
- Oligonucleotide content
- higher-level features of gene
- chromatin structure

Methods

Various classification and statistical approaches are used for analysis of the nucleotide sequence context of mutations hotspots (Rogozin, Babenko, Milanesi, Pavlov 2003 *Brief. Bionform.* 4, 210-227).

DNA polymerase η mutation hotspots in *lacZ*

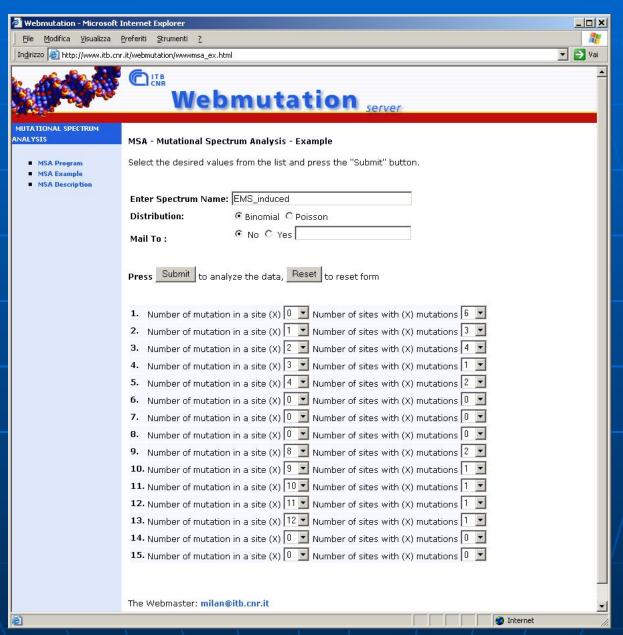
Sequence	Hotspot	Type of	Number of
	position	changes	mutations
CA <u>A</u> TT	3	$A \rightarrow G, T, C$	15,1,1
TT <u>A</u> TC	14	$A \rightarrow G, C, T$	14,1,1
GA <u>A</u> AT	21	A→G,T	16,2
AT <u>A</u> GC	38	$A \rightarrow G, T, C$	9,2,1
CA <u>T</u> AG	39	T→G,A,C	9,9,2
TC <u>A</u> TG	46	A→G,T	13,1
GT <u>A</u> AT	50	A→G,T	16,4
GA <u>A</u> TT	56	A→G	17
AA <u>A</u> CG	70	A→G,T	18,3
GT <u>A</u> AA	73	A→G, T	14,1
CG <u>A</u> CG	80	A→G,T	11,2
\ \			
$W\underline{A}$ Co	onsensus		

Hotspot positions are underlined. Staly Naples

Methods

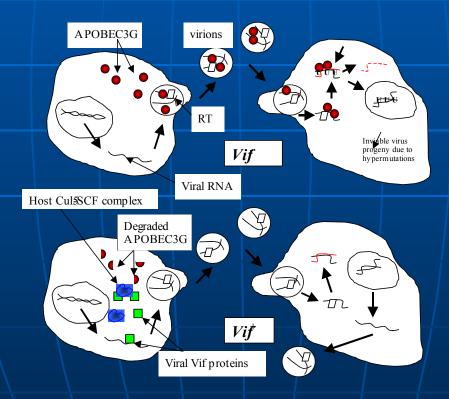
Correlation between nucleotide sequence features and mutation spectra. Nucleotide sequence features (mutable motifs) can be correlated with a mutation spectrum and the correlation can be tested for statistical significance.

								Mo	otifs	
								R <u>G</u> YW/I	WR <u>C</u> Y+W <u>P</u>	$V_{\overline{L}}/V_{\overline{L}}$
		AT						+	-	-
	Γ	AG	С			Mutat	cions	22	7	,
	G	СТ	G							
C	G	TCT	CC			Posit	cions	6	12	2
ATG	г т	CAT A	ACC							
GA <u>T</u>	ATC.	A GC TO	SA <u>TA</u> T(CCA		Fisher	exact	test P	= 0.006	
W.	<u>A</u>	RGYW	WA			(non-ra	indom ta	argeting	of	
<u> T</u> I	W \ 1	WR <u>C</u> Y	<u>T</u> W			mutatio	ns to m	utable :	motifs)	



Hypermutation in HIV-1

The cytidine deaminase APOBEC3G confer resistance to HIV. Its antiviral action could be overcome by the presence of virion infectivity factor (Vif), encoded by the viral genome.



Hypermutation in HIV-1

Spectrum of mutations in the the *GFP* gene induced by the over-expression of APOBEC3G in *E.coli*

A	AA	AAA	AAA		AAAA	AAAAAA			AAAAAA						
A		AAA	AAAA		A				A					A	
A		A		AA	AAAA	AAAAAA			A					A	
A	A	A	A		AA	A		A	A					A	
A	A	A	A		AAA	A	A	A	A					A	
A	A	A A	A A		AAAA	A	A	A	A 2	A	A		A	A A	
ATG	GTGAG	CAA <mark>G</mark> GG	CGAG	AGCTGTTCA	ACC <u>GGG</u> GT	GGTGCCCA!	CCT <mark>GG</mark> TC	:GZ	AGCT <mark>GG</mark> AC	GCGACGTG	AAC <mark>GG</mark> CCA	CAAGTTCAG	CGTGTCC <mark>G</mark> G	CGAGG	100
								,							

GG mutable motif



Spectrum of mutations in HIV-1 DNA in the absence of the Vif protein





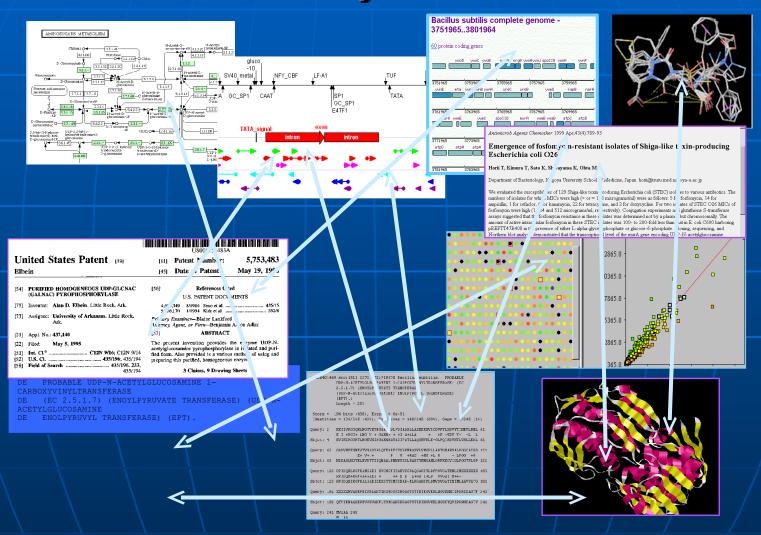
Hypermutation in HIV-1

Specificity of APOBEC3G for <u>G</u>G sequences, which is frequently a part of TGG tryptophan codons, results in a frequent generation of TAG nonsense codons which leads to a premature termination of protein synthesis.

The cytidine deaminase APOBEC3G may cause mutations in HIV-1



HTS Data Project annotations



PROCESSING, ASSEMBLY AND ANNOTATION PROTOCOL

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DB contents and help

A fully automated pipeline has been prepared to process EST sequences using public software integrated by in-house developed Perl scripts.

Sequence files, produced by Dr. Pozzi's lab (libraries S3 and S4), together with the quality files produced by the base-calling program **phred** (Ewing et al, 1998) have been processed in order to identify vector contamination and low quality regions with the program **Lucy** (Chou et al, 2001) using default parameters.

Vector-free high quality sequences have been added to peach EST sequences produced by the groups of Padua (libraries Pp-S4 and S3II) and by the university of Clemson (library PP_Lea) and submitted to the program CAP3 (Huan et al, 1999) to perform contig assembly. Stringency parameters have been modified (-p 95, -d 60) to identify paralogs.

All the input EST sequences and all the contig consensus sequences have been submitted to the **BLASTx** program for annotation (Altschul et al, 1990). BLASTx compares the six-frame conceptual translation products of a nucleotide query sequence against a protein sequence database. BLASTx is run locally against the Genbank nr protein database (nr contains all non-redundant GenBank CDS translations + RefSeq Proteins + PDB + SwissProt + PIR + PRF). Blast output has been parsed and stored in a MySQL database together with other data produced in the intermediate steps of the pipeline.

The ESTree DB Unigene dataset is defined as the collection of all the non-redundant sequences present in the DB and is derived from CAP3 output marking as Unigene all the singleton sequences and the longest sequence of each contig.

SNP detection has been performed with the program AutoSNP version 9 (Baker et al, 2003). Tgicl parameters have been set to -p 95, -l 60, -v 20 for appropriate EST clustering.

A php-based web interface has been prepared to query the database. Users can view sequence data, BLAST outputs, contig alignments, and global statistics on sequences.

Single sequence and contig report pages are available, and fasta formatted sequences can be copied from here. Sequence quality files are available upon request.

A **text search** utility is available and queries can be performed on the **sequence** and **contig** report tables. BLAST **E-value** intervals can also be selected by users.

Local BLASTn, BLASTp, BLASTx, tBLASTn and tBLASTx programs are also available to perform BLAST and batch BLAST searches on the ESTree database.

In the web pages that present data in tabular form an EXCEL **spreadsheet** view is available, to retrieve and copy data in EXCEL format.

In the SNP report page SNP data can be accessed by sequence GI number, by sequence name or by SNP number.

Complete sequence and contig download is allowed from the Download page both in multifasta , CSV and NCBI format.





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Libraries and

PP Lea

Pp S4

SSII

S3 S4

DB contents and help

developmental stages:

CONTIG

Contig name: Contig19 Contig length: 1311

Sequences in this contig: Pp-S4_EST1698, PP_LEa0030J08f, PP_LEa0011B19f, PP_LEa0021I03f, S410N4, S319G11,

PP_LEa0028N23f, PP_S3IIsel_A2_C10, S37D16, S411L16, S311K5, PP_S3IIall_A6_C05

Blast Output

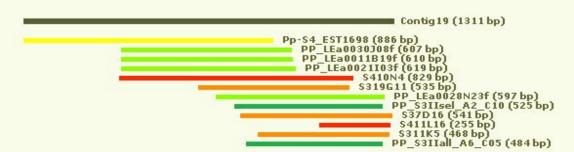
Best Blast Hit: BAD33762.1

E-value: 1e-102

Annotation: putative 6-phosphogluconolactonase [Oryza sativa (japonica cultivar-group)]

Source: Oryza sativa (japonica cultivar-group)...

View alignment details



>Contig19 length=1311 ATCGGTCGCGCTACNTCTCGCGCGCCGTTGATCTCACTCCGAGTGTCACCAATATTCCAA GCTCTCATCGATTTAAATCCGTTCGCACCAATTTCAAGAACCCAATTTCTGCATCGGCAT CAGCATCAGCTAGGATGGCGGGTCAGAATAAGAAGATAGAGAAGTTTGAGACGGAGGAGG AAGTGGCGGTGCGTTTGGCCAAGTACACCGCAGATCTGTCCGCTAAGTTNGTGAAAGAGA





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Welcome to ESTuber

A data bank of Est sequences developed at the Istituto Biologia e Biotecnologia Agraria in the frame of the CNR Strategic Project TUBER: biotecnologia della micorrizazione.

The ESTuber Consortium:

ESTuber is a Consortium of several research centers in Italy devoted to the implementation of genomics and functional genomics in truffle species.

The primary objectives of the Consortium are:

the development of an extensive EST database for Tuber species (ESTuber DB); the analysis of several biochemical pathways based on oligonucleotide microarrays derived from ESTs collection; the analysis of conserved genetic modules in truffle species and related filamentous fungi.

The current members of the Consortium are:

Istituto Biologia e Biotecnologia Agraria, IBBA-CNR, Milano
Istituto Genetica Vegetale, IGV-CNR, Section Perugia
Istituto per la Protezione delle Piante, IPP- CNR, Section Torino
Istituto di Biologia e Patologia Molecolari, IBPM-CNR, Roma
Dipartimento di Biochimica e Biologia Molecolare Università degli Studi di Parma
Istituto di Chimica Biologica, Centro di Biochimica delle Proteine, Università di Urbino

The ESTuber DB:

The ESTuber DB is a collection of Tuber borchii EST sequences.

The ESTuber DB has been developed by:

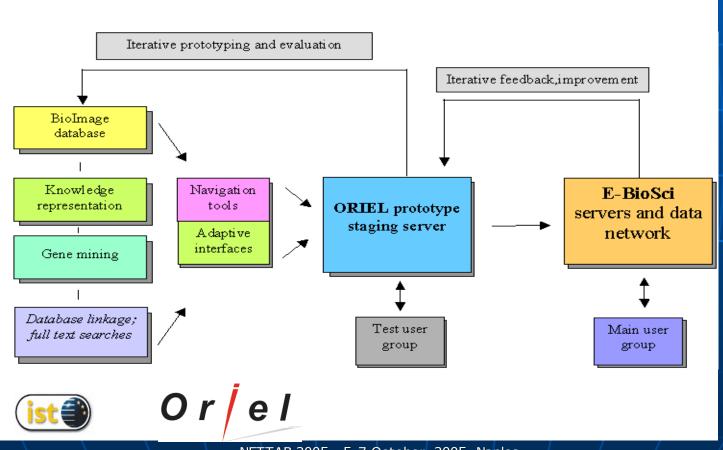
Dr. Barbara Lazzari and Dr. Andrea Caprera: DB designers and webmasters

Dr. Cristian Cosentino and Dr. Barbara Lazzari: cDNA library production and sequences managment

Dr. Luciano Milanesi: Bioinformatics

Dr. Angelo Viotti: ESTuber project manager

ORIEL & E-BIOSCI



GRID Networked data

Laboratory for Interdisciplinary Technologies in Bioinformatics

The GRID: networked data processing centres and "middleware" software as the "glue" of resources.



Researchers
perform their
activities
regardless
geographical
location,
interact with
colleagues, share
and access data

Scientific instruments and experiments provide huge amount of data

NETTAB 2005 5-7 October, 2005 Naples Italy

LITBIO Laboratory for Interdisciplinary Technologies in BIOinformatics



http://www.litbio.org

Molecular medicine has progressed to the point where the majority of human genes and proteins have been characterized and computational methods are necessary for further understanding of structure, mechanism and function.

Our aim is to develop a Laboratory for Interdisciplinary Technologies in Bioinformatics (LITBIO) applied to Genomics, Transcriptomics, Proteomics and Metabolomics.